



## El Colegio de la Frontera Sur

La selección como estrategia para mejorar la competitividad sexual de los machos de cría masiva de *Anastrepha ludens* (Díptera: Tephritidae).

TESIS

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por

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## **Dedicatoria**

A la vida, que ha sido benevolente conmigo y me permite la dicha de concluir esta etapa junto a las personas que más amo.

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Por sus palabras de aliento, por su apoyo y comprensión, por creer que juntos podemos con esto y más.

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## I. Introducción

Las moscas de la fruta (Diptera: Tephritidae) constituyen una de las plagas de mayor importancia en todo el mundo debido a su impacto económico directo y a las estrictas restricciones cuarentenarias impuestas por muchos países (Aluja y Mangan 2008). Las especies que mayor atención han recibido por el impacto negativo que ocasionan en la producción y comercialización de frutos, se encuentran en los géneros *Anastrepha*, *Ragoletis*, *Batrocera*, *Toxotrypana*, *Dacus* y *Ceratitis* (Hernández-Ortiz y Aluja 1993).

El género *Anastrepha* (Schiner) es considerado el grupo más diverso de la familia Tephritidae y el de mayor importancia en América (Hernández-Ortiz et al. 2010). En México se reportan 37 especies del género *Anastrepha*, de las cuales solamente cuatro son de importancia agrícola: *Anastrepha ludens*, *A. obliqua*, *A. striata* y *A. serpentina* (Hernández-Ortiz 2007). La mosca Mexicana de la fruta, *Anastrepha ludens* (Loew), es considerada una de las plagas más severas tanto de los cítricos (*Citrus spp.*) como del mango (Aluja et al. 2014).

Debido a su importancia, en 1992 las moscas de la fruta fueron declaradas plagas de interés público (Rull et al. 1996, Reyes et al. 2000) por lo que se implementó la Campaña Nacional Contra Moscas de la Fruta (CNCMF) (NOM-023-FITO-1995, NOM-075-FITO-1997). Esta Campaña aplica un manejo integrado de plagas con enfoque en áreas grandes donde la Técnica del Insecto Estéril (TIE) ha jugado un papel clave, logrando establecer zonas libres y/o de áreas de baja prevalencia de moscas de la fruta en el norte del país (Hendrichs et al. 2007, Liedo y Toledo 2007, Gutiérrez y Santiago 2008, Gutiérrez 2010).

La TIE es un método de control de plagas amigable con el ambiente, ampliamente utilizado en diversas partes del mundo, y de gran relevancia en el control de varias especies de moscas de la fruta donde se ha aplicado con éxito (Enkerlin 2005). La aplicación de esta técnica requiere de la cría, esterilización y liberación sistemática de un gran número insectos de la especie plaga, con la finalidad de que los machos estériles compitan exitosamente con los machos silvestres por aparearse con hembras silvestres. Por lo tanto, la eficiencia de la TIE depende críticamente de la capacidad de los machos estériles para copular a las hembras silvestres, induciendo esterilidad y en última instancia reducir la tasa de crecimiento poblacional de la plaga (Knipling 1955, Hendrichs et al. 2002, Liedo et al. 2010).

Los insectos de cría masiva experimentan ambientes bióticos y abióticos muy diferentes de aquellos en los que se desarrollan los insectos silvestres. Estas diferencias ambientales tales como la densidad, la dieta artificial de las larvas y adultos, intensidad de luz entre otras, pueden afectar el desempeño de los machos e influir en su capacidad de apareamiento (Cayol 2000, Lux et al. 2002, Lance y McInnis 2005, Rull et al. 2005). Lo anterior ha propiciado que diversas líneas de investigación se orienten al desarrollado de estrategias que permitan equilibrar los efectos negativos de la cría masiva sin comprometer los atributos favorables de la producción masiva (Cayol 2000, Sørensen et al. 2012). Algunos de los métodos implementados para mejorar el desempeño de los machos de cría masiva se han basado en uso de mejores dietas, uso de hormonas, aromaterapia, desarrollo de cepas de sexado genético, manejo de las colonias madre,

modificaciones enfocadas al proceso de colonización, entre otras (Robinson y Hendrichs 2005, Liedo et al. 2007, Pereira et al. 2013).

Por otro lado, las investigaciones orientadas a la ecología evolutiva del comportamiento de moscas de la fruta han permitido desentrañar la dinámica de elección de pareja (Burk 1981, Calkins 1984, Arita y Kaneshiro 1989, Whittier et al. 1995, Eberhard 1996, Benelli et al. 2012), lo que contribuye al conocimiento sobre los rasgos y aptitudes primordiales para el éxito reproductivo de los machos estériles. Por tal motivo, la selección artificial de los rasgos o atributos considerados como buenos indicadores de la aptitud del macho representan una promisorio alternativa para mejorar la eficiencia de la TIE (Whittier et al. 1995, Boake 1996, Benelli et al. 2014).

Estudios relacionados con la selección de machos por su propensión a copular mostraron ser efectivos (Harris et al. 1986, 1988). Sin embargo, estos fueron realizados en condiciones de laboratorio. El conocimiento sobre el sistema de apareamiento lek-polígamo de varias especies de moscas de la fruta (Prokopy y Hendrichs 1979, Burk 1981) mostró que la mayor propensión a copular en condiciones de laboratorio estaba asociada a una menor selectividad de las hembras y por consecuencia un cortejo menos elaborado por parte de los machos. Esto resultó que bajo condiciones de campo las hembras silvestres rechazaban a esos machos (Calkins et al 1984). Estudios más recientes han documentado una mejora en el desempeño de los machos de cría masiva mediante distintos sistemas de manejo de colonia. Rull y Barreda-Landa (2007) demostraron que es posible mejorar el desempeño de los machos estériles de *A. ludens* mediante la hibridación de machos silvestres con hembras de cría masiva. McInnis et al. (2002) mejoraron el desempeño de los machos estériles de *Ceratitis capitata*, a través



de un proceso combinado de selección e hibridación. Aunque estos estudios han confirmado que es posible mejorar el desempeño de los machos de laboratorio mediante cruzamientos con cepas silvestres o con determinadas características deseables, los procesos de hibridación representan dificultades para su implementación en la cría masiva y pueden resultar complicados cuando se crían cepas de sexado genético o genéticamente modificadas. Ante estas dificultades, la selección de genotipos con alto éxito reproductivo adquiere mayor relevancia. Recientemente se ha comprobado el efecto positivo de la selección en *A. ludens* (Bosa et al. 2016, Quintero et al. 2016).

Aunque los resultados de la selección son promisorios, aun se requiere fortalecer el conocimiento respecto a los efectos de los procesos de selección en múltiples atributos biológicos clave para la producción masiva. Debido a que la selección representa una posibilidad para mejorar el desempeño de los insectos estériles y por lo tanto la efectividad de la TIE, el presente trabajo tuvo como objetivo comparar el efecto de un proceso de selección en parentales, y de selección continúa a lo largo de cuatro generaciones, en la competitividad sexual de machos de *A. ludens* y su efecto en algunos atributos biológicos (sobrevivencia y reproducción).

En el siguiente capítulo se presentan los resultados de esta investigación de acuerdo al manuscrito que fue sometido para publicación en la revista *Entomologia Experimentalis et Applicata*. En el capítulo III se presenta una discusión general y las principales conclusiones del trabajo y en el capítulo IV la literatura citada en esta introducción y en el capítulo III.

**II. Effect of artificial selection on mating competitiveness and demography of *Anastrepha ludens* (Diptera: Tephritidae) for sterile insect technique application**

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20 **Effect of artificial selection on mating competitiveness and demography of**  
21 ***Anastrepha ludens* (Diptera: Tephritidae) for sterile insect technique application**

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36 **Running head:** Selection on mating competitiveness for SIT

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## Abstract

The sterile insect technique (SIT) has been applied successfully for the control of several fruit fly species of economic importance. However, mass-rearing conditions have resulted in selection of sexual behavioral traits that differ from wild flies reducing mating competitiveness of sterile flies and SIT efficiency. Artificial selection in mass-rearing colonies represents an alternative to improve the sexual performance of sterile flies. Here, we evaluated the effect of selection of *Anastrepha ludens* (Loew) (Diptera: Tephritidae) mass-reared males based on their mating competitiveness. Two modes of selection were compared, one single selection event on parental flies, and continuous selection along four consecutive generations. Field cage mating tests showed that wild males were more competitive than mass-reared males. Although no significant differences were found between mass-reared selected and non-selected males, in the fourth generation, males from the selected colonies performed better and were closer to the wild males than males from the non-selected colonies. Mating latency tended to be longer for wild males, and males from selected colonies tended to take longer to mate, but significant differences were only found in the second generation. No consistent differences or patterns were observed in copula duration or female mating inhibition. Wild females had greater life expectancy and lower reproductive rates than mass-reared females. Survival and fecundity decreased with increasing rearing generations, except in the colony with continuous selection. Our results suggest a trade-off between mating competitiveness and survival and reproductive rates. Reasons for disparity in results on selection of sexual performance, and reduction in survival and reproductive rates through rearing generations are discussed.

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## Introduction

The sterile insect technique (SIT) is an environmentally friendly pest control method that has been used successfully against some species of tephritid fruit flies (Enkerlin, 2005; Robinson & Hendrichs, 2005; Gilchrist & Meats, 2014). This technique requires the mass production, sterilization and release of the target species with the aim that sterile males will mate with wild females inducing sterility and reducing the wild population growth rate (Knipling, 1955; Hendrichs et al., 2002; Liedo et al, 2010).

The loss of sexual competitiveness of the mass-reared sterile males could be a limiting factor for the successful application of the SIT (Calkins, 1984). This loss could be attributed to the effects of irradiation and/or the artificial conditions used for mass-rearing that inadvertently affect the physiology and behavior of the sterile insects. The time for initiation of sexual activities, male ability to inhibit female remating, male courtship patterns, and acoustic, visual and chemical signals are examples that affect female mating choice and the effectiveness of SIT (Liedo & Carey, 1996; McInnis et al., 1996; Cayol, 2000; Vera et al., 2003; Lance & McInnis, 2005; Lux et al., 2007ab).

Differences in biological traits of wild and mass-reared insects are attributed to different selection factors under mass-rearing conditions and the natural environment. Mass-reared insects are characterized for their high fecundity and short developmental time (Greanny & Szentesi, 1979; Vargas & Carey, 1989; Meats et al., 2004; Hernández et al., 2009), which are favorable adaptations for mass production. However, differences between the sexual behavior of mass reared flies compared to wild flies can have adverse effects on the performance of sterile insects (McInnis et al., 1996; Lance et al., 2000; Rull et al., 2005; Lux et al., 2007, Matsuyama & Kuba, 2009; Meza et al., 2014). Strategies trying to balance mass-rearing efficiency and field sexual performance

86 have been investigated (Cayol, 2000; Liedo et al., 2007; Sørensen et al., 2012). For example, post-  
87 teneral diets, hormone analogues and aromatherapy have improved the field performance of sterile  
88 males (Pereira et al., 2013).

89 Colonization process or colony management methods have been proposed to improve the mating  
90 competitiveness of sterile insects under natural conditions. Rull & Barreda-Landa (2007) showed  
91 that it was feasible to improve the performance of *Anastrepha ludens* (Loew) sterile males through  
92 hybridization of wild males with mass-reared adapted females, without affecting the productivity  
93 required for mass production. Gilchrist & Meats (2014) found that outcrossing *Bactocera tryoni*  
94 (Froggatt) strains partially adapted to laboratory conditions with the mass-reared adapted strain  
95 produced heat tolerant flies with high egg production. McInnis et al. (2002) were able to improve  
96 the performance of mass-reared *Ceratitidis capitata* (Wiedemann) males through a combination of  
97 selection and hybridization. However, hybridization process might be complicated to implement  
98 in mass-rearing protocols, particularly when genetic sexing strains or genetic modified strains are  
99 produced.

100 The Mexican fruit fly, *Anastrepha ludens*, is a major pest of some citrus fruits (*Citrus* spp.) and  
101 mango (*Mangifera indica* L.) in Mexico and Central America (Aluja, 1994; Thomas, 2003). As  
102 part of a national area-wide integrated pest management program, the use of the SIT to control this  
103 pest species has been implemented. Our aim here was to study the effect of artificial selection  
104 based on sexual competitiveness on the mating performance of mass-reared flies and their life  
105 history traits. Two modes of selection were compared, selection on the parental generation only,  
106 and selection in each generation through four consecutive generations.

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## Materials and Methods

### *Study site*

The study was carried out under laboratory and field cage conditions. Standard *A. ludens* rearing and experiments to test mating inhibition and determination of demographic parameters were done under laboratory conditions at El Colegio de la Frontera Sur, in Tapachula, Chiapas, México. Laboratory conditions were 25±2°C, temperature, 65±5% relative humidity, and 12L:12D photoperiod. The selection process and tests to assess mating competitiveness were done in field cages located at the experimental orchard of the Moscafrut facility (SAGARPA-IICA) located in Metapa, Chiapas (14°49'33.8"N 92°11'46.1"W), and at 100 m above sea level (masl). Environmental conditions were 25-30°C temperature, 70-90% relative humidity and 3,500-0 Lux light intensity. Field cages were 3 m in diameter by 2 m high (Calkins & Webb, 1983) with a ca. 1.8 m in height mango tree placed inside, at the center of the cage.

### *Insects*

Parental mass-reared flies were obtained as pupae from a production batch selected at random from the standard production at the Moscafrut facility with approximately 80 generations. Wild flies were obtained as larvae from infested sour oranges (*Citrus aurantium* L.), grapefruits (*C. paradise* L.) and white sapote or matasano (*Casimiroa edulis* Llave & Lex) collected in the Soconusco region and in the Comiteca Tojolabal plateau, Chiapas, México. Infested fruits were brought to the laboratory and placed in plastic trays (60x50x10 cm) with vermiculite. Once third instar larvae were mature, they were extracted from the fruits and placed in 27x16x4 cm containers with vermiculite at 60% humidity, to promote pupation.

131 At eclosion, adult flies were sorted by sex and placed in glass cages (30x30x30 cm) with water  
132 and standard laboratory diet (3:1 sugar: yeast hydrolyzate enzymatic, [ICN Biomedicals, Costa  
133 Mesa, California, USA]). Mass-reared and wild flies were maintained under these conditions until  
134 they reached sexual maturity (~14 days for mass-reared flies and ~19 days for wild flies). Wild  
135 flies were collected for each test in each generation.

136

### 137 *Rearing process*

138 Standard rearing processes were followed for each treatment and generation. Adults were  
139 maintained in 30x30x40 cm cages. At the top of the cage, a 16 cm in diameter egg chamber  
140 was placed. This was made of black tergal cloth coated with silicon, containing water to avoid egg  
141 desiccation. The eggs laid in the chamber were placed over wet black satin cloth in a 19 cm in  
142 diameter Petri dish. After ~4-5 days the larvae hatched and were transferred to the artificial larval  
143 diet in plastic trays at a density of 5 larvae per gram of diet (Domínguez et al., 2010).  
144 Approximately 10 day later, when the larvae were mature and ready to pupate, they were sorted  
145 out from the diet and placed in 27x16x4 cm containers with vermiculite at 60% humidity to  
146 promote pupation. After 10 days, pupal size was standardized by means of a pupa sorter machine  
147 with 10 size categories (FAO/IAEA/USDA, 2003). Based on size frequency distribution, the most  
148 frequent categories, 7 (2.30-2.45 mm in diameter), 8 (2.45-2.60 mm in diameter) and 9 (2.60-2.75  
149 mm in diameter), were used. Fourteen days after pupation, pupae were sorted out from the  
150 vermiculite and placed in 30x30x30 cm glass cages for adult emergence. At emergence, adults  
151 were sorted by sex and maintained in independent cages, at similar densities (200 – 250 flies per  
152 cage), provided with water and food as described above, until they reached sexual maturity.



153 *Treatments*

154 Three colonies were maintained under the previously described standard rearing process:

155 1) Laboratory (L). This was a control colony with no selection applied. Parental flies were taken  
156 as a random sample from the standard rearing at the Moscafrut facility.

157 2) Alpha Selection (AS). This was the colony based on only a single selection event on the parental  
158 flies. The colony was started with laboratory males that successfully mated with wild females, in  
159 competition with wild males. For the selection process, 100 laboratory males, 100 wild males, and  
160 100 females were released in each of 8 field cages. Laboratory males were 14 days-old and wild  
161 flies were 19 – 23 days old. Laboratory males that successfully mated (intromission longer than  
162 one minute) were selected. The process was repeated the following day, using the selected males  
163 and maintaining the 1:1 laboratory: wild male ratio and the 2:1 male: female ratio. The process  
164 was repeated until selected males accounted for ~1% of the initial cohort (800 males). This  
165 normally occurred after 3 consecutive days. Selected males were placed in reproduction cages with  
166 water and standard food under laboratory conditions. Then, laboratory sexually mature females  
167 were introduced in these cages at a 3:1 female: male ratio. The same number of females was added  
168 every 3 days for 12 days to assure enough offspring of the F1 generation from the selected males.

169 3) Beta Selection (BS). In this colony, the selection process was repeated in each generation during  
170 3 more generations (until F4). The field cage tests used to assess mating performance was the  
171 initial step for the selection process. AS males that successfully mated with wild females were  
172 selected and tested again the next day. The process was repeated until ~1% of the initial cohort  
173 was selected. In the next generations, BS males that successfully mated were selected following  
174 the same procedure and their offspring were obtained as described above. Figure 1 graphically

175 describes the process and indicates the precise number of insects used in the field cage tests and  
176 the number and percent of selected males in each generation.

177

### 178 *Mating competitiveness*

179 Virgin males, age 14-17 days old of each of the three colonies (L, AS, BS) and 19-23 days old  
180 wild males and females were used in the field cage tests. Five days before the test, males were  
181 marked with a small dot of acrylic paint (Baco® Mexico City) on the thorax, using different colors  
182 to identify each treatment. Marked males were maintained in the laboratory in 15x15x25cm cages,  
183 provided with water and standard food.

184 Tests started 2 hours before sunset and finished just after sunset, when it was completely dark. In  
185 each field cage, 75 to 100 males of each colony (L, AS, BS) were released together with 75 to 100  
186 wild males. Twenty minutes later, 150 wild females were released into the cage. Mating pairs were  
187 collected in vials and the time at mating and mating duration were recorded. Mating latency was  
188 defined as the time from the release of the females in the field cage to the time of the first mating.  
189 Mating duration was defined as the time each pair remained in copula.

190 Each cage was considered as a replicate. The number of replicates were 4, 6, 3 and 3 for F1, F2,  
191 F3 and F4 generations, respectively. The density of flies per cage varied according to the number  
192 of treatments evaluated. In the F1 generation only L and AS males were tested, BS males were  
193 added in the next 3 generations. The 1:1 male: male ratio and the 2:1 male: female ratio were  
194 constant in the four generations tested (Figure 1).

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198 *Mating inhibition*

199 Wild females that mated with males from the different treatments were placed individually in 7  
200 cm diameter by 10 cm long labeled containers, provided with water and standard diet, under  
201 laboratory conditions. Three days later a 15-22 days old laboratory virgin male was introduced in  
202 each container. Individuals were observed during 2 hours and the number of rematings were  
203 recorded according to the type of male (L, AS, BS, W) females had initially mated with. The  
204 proportion of females remating after 3 days from the first mating was used as an indicator of the  
205 initial male's ability to inhibit female remating.

206

207 *Demographic parameters*

208 Immature and adult survival, fecundity and fertility of the offspring of each treatment (L, AS, BS)  
209 and each generation were estimated to see if selection based on sexual competitiveness was related  
210 to these biological attributes.

211 Immature survival was estimated from an initial sample of 100 eggs for each colony, obtained  
212 from the egg chambers described above. These eggs were placed on black satin fabric placed  
213 over a moistened sponge in plastic Petri dishes (9 cm diameter). These dishes were observed daily,  
214 for 8 days, under the microscope to determine egg hatch. For larval survival, eggs ready to hatch  
215 were placed on artificial diet (two eggs per gram of diet) formulated according to the Moscafrut  
216 facility mass-rearing process (Domínguez et al., 2010). The diet was placed in plastic Petri dishes  
217 (9 cm diameter) with 50 g of diet. Once the larvae reached the third instar and were ready to pupate,  
218 survival was recorded, and the larvae were placed in 6 cm in diameter by 5 cm high plastic  
219 containers with vermiculite at 60% humidity. Pupal survival was estimated from the proportion of  
220 adults that emerged. Ten replicates were done for each treatment and generation.

221 Adult survival, fecundity and fertility were estimated from samples of 50 flies, 25 males and 25  
222 females for each treatment and each generation. At eclosion, individual pairs (male and female)  
223 were placed in 7 cm in diameter by 10 cm high containers provided with water and standard diet.  
224 The number and sex of dead individuals was recorded daily, until the last individual died.  
225 Each container had an egg device on top. The egg device consisted of a plastic cup that fit  
226 into the mouth of the bottle (6 cm in diameter by 3 cm high). The bottom of the cup was replaced  
227 with black satin fabric coated with silicon and the cup was filled with water. Females laid their  
228 eggs through the silicon-coated fabric and the eggs were collected daily from the water with a  
229 pipette. The eggs of each individual female were placed on black satin fabric placed over a  
230 moistened sponge in a plastic Petri dish. Fecundity was estimated from the number of eggs laid  
231 per female per day. Fertility was considered as the number of eggs that hatched (fertile eggs).  
232 Collected eggs were observed under the microscope six days later to record the number of hatched  
233 eggs.

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#### 235 *Statistical analysis*

236 For the mating competitiveness test, the number of matings by each treatment in each field cage  
237 were transformed to proportions. These data were subject to one-way analysis of variance  
238 (ANOVA). In generations where the ANOVA assumptions of normality and homoscedasticity did  
239 not fit, Box-Cox transformations were used. When differences among treatments were significant  
240 ( $P < 0.05$ ) in each generation, a Tukey HSD (Honestly Significant Different) test was done to  
241 compare means.

242 Mating latency and copula duration data were adjusted to a linear mixed-effects model. This was  
243 because of the interactions among flies in the field cage and the one level hierarchy, where the

244 cage was considered the random effect and the treatments were considered the fixed effects. When  
245 differences were significant ( $P < 0.05$ ), a Tukey multiple means comparison was made.

246 Mating proportions of each treatment through the 3 generations were analyzed as a split-plot  
247 design to determine if there were interactions between treatments (L, W, AS, BS) and generations  
248 (F2, F3, F4). Tukey multiple means comparisons were made where differences were significant.

249 This analysis was made only for the last 3 generations, because in the F1 generation the BS  
250 treatment was absent. To balance the number of replicates, in F2 generation 3 replicates were  
251 selected at random.

252 Mating inhibition was measured as the proportion of wild females that remated. Contingency  
253 tables were used for the analysis on the condition of the female (remated / did not remate), and the  
254 male treatment of the initial mating (W, L, AS or BS). In each of the generations where significant  
255 difference was found, associations between factors were verified by calculating the proportion of  
256 contribution of each cell in the table in relation to the value of  $\chi^2$  (Zar, 2010).

257 Immature survival was considered as the number of individuals that survived to the next stage.  
258 Data were analyzed by a linear regression model adjusted to generalized least squares, to allow for  
259 variance heterogeneity and residuals dependency. In generations where significant differences  
260 were found, Tukey multiple means comparisons were made.

261 Adult demographic parameters were estimated following methods described by Carey (1993).  
262 Survival curves were analyzed by Log-rank test for interval censored data considering that  
263 mortality was recorded every 24 hours (Therneau & Grambsch, 2000; Fay & Shaw, 2010). Gross  
264 fecundity ( $\sum m_x$ ) was analyzed by ANOVA, followed by HSD or Fisher LSD test. Fertility rates  
265 were analyzed by binomial logistic regression and Tukey multiple mean comparison test.

266 Analyses were carried out using R software version 3.2.4 (R Core Team, 2016) and the packages  
267 Icats (Gentleman & Vandal, 2016) MLEcens (Maathuis, 2013) and nlme (Pinhero et al., 2016).

## 268 **Results**

269

### 270 *Mating competitiveness*

271 Wild males (W) were the most successful in terms of the proportion of matings obtained in each  
272 of the four generations evaluated (Figure 2). However, differences were significant only for the  
273 first 3 generations (F1:  $F_{2,9} = 39.449$ ,  $P = 3.517e-05$ ; F2:  $F_{3,20} = 4.3329$ ,  $P = 0.0165$ ; F3:  $F_{3,8} =$   
274  $19.52$ ,  $P < 0.0004$ ; and F4:  $F_{3,8} = 2.969$ ,  $P = 0.0971$ ). No significant differences were observed  
275 among AS, BS and L males, although in the F4, the selected colonies performed better and were  
276 closer to the wild males than the L males. There were not significant differences among generations  
277 ( $F_{2,6} = 0.456$ ,  $P = 0.654$ ), but differences among treatments were significant ( $F_{3,18} = 13.316$ ,  $P =$   
278  $8.05e-05$ ). The *post hoc* test confirmed that wild males were more successful in mating than males  
279 of the three rearing colonies (L, AS, BS). There was no significant interaction between generations  
280 and treatments ( $F_{6,18} = 2.095$ ,  $P = 0.105$ ).

281

### 282 *Mating latency and copula duration*

283 Mating latency tended to be longer for wild males than for mass-reared males, and selected males  
284 tended to be closer to wild males, but significant differences were only observed in the F2  
285 generation ( $F_{3,209} = 5.7793$ ,  $P = 8e-04$ ) (Table 1). There were no significant differences in copula  
286 duration, except in the F2 generation, where wild males had the longest duration ( $F_{3,209} = 8.0470$ ,  
287  $P < 0.0001$ ).

288

289 *Mating inhibition*

290 The percentage of wild females that remated ranged from 6.3 to 25.5% (Figure 3). In the F1  
291 generation, females that mated with wild males showed the lowest remating percentage, but  
292 differences were not significant ( $\chi^2_2 = 0.1192$ ,  $P = 0.9421$ ). In the F2 generation, again females that  
293 first mated with wild males showed the lowest percentage of remating and in this case differences  
294 were significant ( $\chi^2_3 = 10.779$ ,  $P = 0.0129$ ). Tukey *post hoc* tests revealed significant differences  
295 between females that mated with W males and females that mated with AS males. Differences in  
296 remating frequency were not significant in F3 ( $\chi^2_3 = 1.8517$ ,  $P = 0.6038$ ) and F4 generations ( $\chi^2_3$   
297  $= 2.4911$ ,  $P = 0.4769$ ).

298

299 *Demographic parameters*

300 Immature survival decreased in each generation. There were significant interactions between  
301 treatments and stages of development in F2 and F3 ( $F_{2,83} = 9.2672$ ,  $P = 0.0002$  and  $F_{2,83} = 15.2699$ ,  
302  $P < 0.0001$ , respectively), but not in F1 ( $F_{1,55} = 0.0388$ ,  $P = 0.8445$ ). Survival in the L control was  
303 greater than in the treatments with selection (AS, BS), and was lower in BS for all developmental  
304 stages (Figure 4). According the multiple mean comparisons, in F2, survival in the L control was  
305 greater than in the selection treatments in the 3 developmental stages (larva, pupa and adult). In  
306 F3, larval survival was different among the 3 treatments and L individuals showed the greatest  
307 survival. No differences were found in survival to pupae and adults.

308 Adult demographic parameters are shown in table 2. Flies from the BS colony were evaluated only  
309 in F2 and F3 generations, while the parameters of W flies were from only one cohort. Wild flies  
310 showed greater female life expectancy and lower reproductive rates compared with the three mass-  
311 reared colonies. In mass-reared flies, female life expectancy decreased throughout the rearing

312 generations, with the only exception in the case of the BS colony. There was not a clear trend in  
313 male life expectancy, but the F3 generation showed the lowest values in the three mass-reared  
314 colonies.

315 There were significant differences in male and female survival (Figure 5). In females, differences  
316 were found in the 3 generations (F1:  $\chi^2_2 = 7.9493$ ,  $P = 0.0187$ ; F2:  $\chi^2_3 = 18.7137$ ,  $P = 0.0003$  y  
317 F3:  $\chi^2_3 = 26.8686$ ,  $P = 6.273e-06$ ). According to the post hoc test W females showed greater  
318 survival than females from the mass-reared treatments (L, AS, BS) in the 3 generations, except in  
319 the F2 generation where the difference between W and L was no significant. There were not  
320 significant differences among mass-reared treatments. In males, no significant differences were  
321 found in F1 and F2 generations (F1:  $\chi^2_2 = 3.0935$ ,  $P = 0.2129$ ; F2:  $\chi^2_3 = 2.6082$ ,  $P = 0.4560$ ). In  
322 F3 there were significant differences (F3:  $\chi^2_3 = 15.9752$ ,  $P = 0.0011$ ). According to multiple mean  
323 comparisons, W and L males had greater survival than AS and BS males, with no differences  
324 between L and W or between AS and BS males.

325 Gross and net fecundity rates were greater in the mass-reared colonies than in the wild strain.  
326 However, these rates decreased with rearing generations. The only exception was the BS colony,  
327 where fecundity rates increased from the F2 to the F3 generations (Table 2). In the F1 generation,  
328 gross fecundity of the L colony was significantly greater than the fecundity of the AS colony (F<sub>1</sub>,  
329  $F_{45} = 10.055$ ,  $P = 0.0027$ ). In F2, no significant differences were found (F<sub>2,53</sub> = 1.1822,  $P = 0.3146$ )  
330 and in the F3 generation differences were significant (F<sub>2,52</sub> = 3.4648,  $P = 0.0386$ ). According to  
331 Fisher LSD test, BS flies showed significantly greater fecundity than L and AS flies. Figure 6  
332 shows the gross fecundity schedules for the four treatments in the 3 rearing generations. Wild flies  
333 showed a longer oviposition period, but daily egg production was lower than in the mass-reared  
334 colonies during peak production. The reproductive period for the three mass-reared colonies was



335 reduced in each generation. Regarding fertility, significant differences were observed in the three  
336 generations (F1:  $\chi^2_1 = 215.43$ ,  $P < 2.2e-16$ ; F2:  $\chi^2_2 = 1834.3$ ,  $P < 2.2e-16$  y F3:  $\chi^2_2 = 406.18$ ,  $P <$   
337  $2.2e-16$ ) (Figure 7). Multiple mean comparisons indicated differences among the 3 treatments and  
338 the 3 generations. In F1, L flies showed greater fertility than AS flies. However, in F2 and F3  
339 generations, AS and BS showed the greatest fertility rates, and L the lowest (Figure 7).

## 340 **Discussion**

341 Mating field cage tests confirmed that wild males were more competitive than mass-reared males  
342 throughout the four generations evaluated. This negative effect of mass-rearing conditions on  
343 mating competitiveness has been documented for a number of tephritid species, including *A.*  
344 *ludens* (Briceño et al., 1998; Miyatake, 1998; Lance et al., 2000; Ávila et al., 2011; Meza et al.,  
345 2014).

346 The effect of selection (AS and BS) was not as expected despite the high level of selection pressure  
347 (F1 = 98.5%, F2 = 98.7%, F3 = 95.1%). In the F4 generation more matings were achieved by  
348 males from the AS and BS colonies than males from the L colony and these were closer to the W  
349 males, but differences were not significant. The positive effect of selection on mating  
350 competitiveness was shown by McInnis et al. (2002) in *C. capitata*, and by Bosa et al. (2016) in  
351 *A. ludens* and Quintero et al. (2016) for a genetic sexing strains of *A. ludens*. A possible explanation  
352 for this unexpected result could be attributed to the use of large numbers of L females for  
353 reproduction. These females were used because they were already adapted to mass-rearing  
354 conditions and facilitate the production of large numbers of offspring. However, their genetic  
355 contribution could result in greater variation and no significant effect on their male offspring  
356 mating competitiveness. Indeed, mass-rearing seems to affect more female parameters than male  
357 parameters (Meza et al., 2014).

358 Another possible explanation might be that the initial numbers used in our selection experiments  
359 (800 in the parental generation and 450 to 225 in the subsequent generations) were not large  
360 enough, or that the number of initial males due to the strength of selection (6 to 11) was too small.  
361 Population size is a fundamental aspect in selection experiments, since size is related to genetic  
362 variation, recombination probabilities and genetic drift (Falconer & Mackay, 1996; Conner, 2003;  
363 Jalvingh et al., 2014). Inadvertent selection on other traits, inbreeding and genetic drift could  
364 counteract the effect of selection. Based on previous results (Harris et al., 1986; McInnis et al.,  
365 2002; Rull et al (2015); Bosa et al., 2016; Quintero et al., 2016), the potential of selection in colony  
366 management to improve field performance of sterile males could not be discarded, but questions  
367 regarding the optimal population size and selection strength should be addressed.

368 Mating latency tended to be longer in W males than in mass-reared males, although differences  
369 were only significant in the F2 generation. According to Schutze et al. (2015) in *Bactrocera*  
370 *dorsalis* (Hendel) the differences in mating latency between laboratory and wild flies are closely  
371 associated to light incidence during sunset. Here, although no significant differences were  
372 observed, it is interesting that selected males increased this time to mate, more closely resembling  
373 W male behavior. Shorter mating latency of mass-reared *A. ludens* flies compared with W males  
374 has been previously reported (Meza et al., 2014; Quintero et al., 2016). A longer mating latency  
375 means that males start calling and courting later (Bosa et al., 2016). This can be an adaptation of  
376 W males to reduce the time they are vulnerable to predators. But under our field cage conditions,  
377 the advantage of delaying the time to start calling and courting was not observed because flies  
378 were protected from predators. A longer mating latency could also be due to males performing  
379 more elaborate or longer pre-copulatory courtships, which would coincide with the higher mating  
380 success of W males. If mass-reared strains start calling before W males this could be advantageous

381 and is reflected in a shorter mating latency. However, across generations this did not result in  
382 higher mating success. Only in the second generation, the BS colony tended to increase mating  
383 latency. Mating latency is probably determined by a combination of female and male factors. In  
384 *C. capitata* wild females reject males with short pre-mount courtship (Briceño & Eberhard, 2000),  
385 and in *A. ludens* wild females are also considered choosier than laboratory females (Meza et al.,  
386 2014). The shorter mating latency of mass-reared males could thus be attributed to males being  
387 more aggressive or having shorter courtship sequences than W males. In *C. capitata* pre-mount  
388 courtships of mass-reared males is significantly shorter than those of wild males (Briceño &  
389 Eberhard, 2000). The fact that there was no consistent increase in mating latency across  
390 generations suggests again that the female component in determining latency to mate is strong.  
391 No pattern was identified in copula duration, despite the significant differences found in the F2  
392 generation. Apparently, these values could be completely random and not be a good measure of  
393 sexual competitiveness. Pérez-Staples et al. (2010) proposed that copula duration is controlled by  
394 the female and it is not a good indicator of male performance.

395 Also, no consistent differences or patterns were found in mating inhibition that could establish a  
396 relationship between the type of male in the first copula and remating propensity. Our results  
397 suggest that mass-reared males have the same potential as wild males to inhibit remating (Abraham  
398 et al., 2012, 2014; Meza et al., 2014). This could be interpreted as a beneficial trait for SIT, but the  
399 effect of irradiation for sterilization should be taken into account.

400 Immatures survival suggests that L flies are well adapted to the larval diet and laboratory  
401 conditions and selection on mating competitiveness had a negative effect on survival. This could  
402 be a trade-off between these traits, as was reported for *B. cucurbitae* developmental time and  
403 sexual performance (Miyatake 1995). The low survival from egg to first instar larva in the BS

404 treatment could be attributed to the large female: male ratio that resulted in non-fertilized eggs.  
405 Further research, trying to understand the relationship between survival of immatures and sexual  
406 performance could be worthwhile.

407 Greater life expectancy of wild females, compared with mass-reared strains, has been reported for  
408 several tephritid species (Liedo & Carey, 1996). This difference could be explained by the cost of  
409 reproduction. Mass-reared females produce more eggs in a shorter time requiring the depletion of  
410 reserves with consequent shorter life expectancy (Carey et al., 2008). Male life expectancy was  
411 greater than female's and no difference or pattern was detected among treatments. The gradual  
412 decrease of survival with generations was a common feature in all rearing colonies. This could be  
413 attributed to the rearing conditions. To obtain enough eggs and to avoid selection for fast  
414 development, eggs from females of different ages were mixed. This probably resulted in  
415 inadvertent negative selection on adult survival.

416 Reduction in reproductive rates through the rearing generations was unexpected. Almost all studies  
417 comparing wild and mass-reared flies indicate that in mass-reared adapted strains, developmental  
418 time is shorter and fecundity rates are greater (Liedo et al., 1992, 2007; Vargas et al., 2000; Meats  
419 et al., 2004; Ekesy et al., 2007; Hernández et al., 2009). This could also be explained by the  
420 inadvertent selection resulting from our rearing conditions. The reduction in reproductive rates  
421 was a combination of a narrower reproductive window, a decrease in the number of eggs per day  
422 during the peak oviposition period and the decrease in female survival. The BS colony was an  
423 exception. In this case, female life expectancy and reproductive rates significantly increased from  
424 one generation (F2) to the next (F3). This suggests that if this selection process continues for more  
425 generations, fecundity rates could reach the level of the L control.

426 Fertility rates increased in the colonies where selection was applied in the F2 and F3 generations.  
427 This was an unexpected result, but in this case, it is favorable from the perspective of mass-rearing  
428 to SIT application.  
429 Our results suggest that selection on mating competitiveness might have a trade-off with other  
430 biological attributes, such as survival and reproduction. Perhaps these traits are polygenic and the  
431 selection could have resulted in epistatic and/or pleiotropic process (Carey et al., 2005; Jalvingh  
432 et al., 2014; Miyatake, 1998; Rull et al., 2015). Despite the unexpected changes in survival and  
433 reproduction through rearing generations, the differences between wild and mass-reared flies were  
434 sustained. This suggests that adaptation to rearing conditions is well fixed. If mating  
435 competitiveness and other traits related to field performance (i.e. predator avoidance, tolerance to  
436 extreme environmental conditions) could be improved by selection, this will represent a favorable  
437 contribution to more efficient SIT. Understanding the genetic basis of complex polygenetic traits  
438 and their trade-offs, as proposed by Benelli et al. (2014), could contribute to the selection of  
439 genotypes adapted to both, mass-rearing and field conditions.

440

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448

449 **References cited**

- 450 Abraham S, Cladera J, Goane L & Vera MT (2012) Factors affecting *Anastrepha fraterculus*  
451 female receptivity modulation by accessory gland products. *Journal of Insect Physiology*  
452 58: 1–6.
- 453 Abraham S, Nuñez-Beverido N, Contreras-Navarro Y & Pérez-Staples D (2014) Female  
454 receptivity in *Anastrepha ludens* (Diptera: Tephritidae) is not modulated by male accessory  
455 gland products. *Journal of Insect Physiology* 70: 41–48.
- 456 Aluja M, (1994) Bionomics and management of *Anastrepha*. *Annual Review of Entomology*  
457 39:155-178.
- 458 Avila FW, Sirot LK, LaFlamme BA, Rubinstein CD & Wolfner MF (2011) Insect seminal fluid  
459 proteins: identification and function. *Annual Review of Entomology* 56: 21–40.
- 460 Benelli G, Giunti G, Canale A, Messing RH (2014) Lek dynamics and cues evoking mating  
461 behavior in tephritid flies infesting soft fruits: implications for behavior-based control  
462 tools. *Applied Entomology and Zoology* 49:363–373
- 463 Bosa CF, Cruz-Lopez L, Zepeda-Cisneros CS, Valle-Mora J, Guillén-Navarro K & Liedo P (2016)  
464 Sexual behavior and male volatile compounds in wild and mass-reared strains of the  
465 Mexican fruit fly *Anastrepha ludens* (Diptera: Tephritidae) held under different colony  
466 management regimes. *Insect Science* 23: 106-116.
- 467 Briceño RD & Eberhard WG (1998) Medfly courtship duration: a sexually selected reaction norm  
468 changed by crowding. *Ethology, Ecology and Evolution* 10: 369–382.
- 469 Briceño RD & Eberhard WG (2000) Possible Fisherian changes in female mate-choice criteria in  
470 a mass-reared strain of *Ceratitis capitata* (Diptera: Tephritidae). *Annals of the*  
471 *Entomological Society of America* 93: 343–345.

- 472 Calkins CO (1984) The importance of understanding fruit fly mating behavior in sterile male release  
473 programs (Diptera: Tephritidae). *Folia Entomologica Mexicana* 61: 205–213
- 474 Calkins CO & Webb JC (1983) A cage and support framework for behavioral tests of fruit flies in  
475 the field. *Florida Entomologist* 66: 512–514.
- 476 Carey JR (1993) *Applied Demography for Biologist with Special Emphasis on Insects*. Oxford  
477 University Press. New York. 206 pp.
- 478 Carey JR, Liedo P, Müller HG, Wang JL, Senturk D & Harshman L (2005a) Biodemography of a  
479 long-lived tephritid: Reproduction and longevity in a large cohort of Mexican fruit flies,  
480 *Anastrepha ludens*. *Experimental Gerontology* 40:793–800.
- 481 Carey JR, Harshman L, Liedo P, Müller HG, Wang JL & Zhang Z (2008) Longevity-Fertility  
482 Trade-offs in the Tephritid Fruit Fly, *Anastrepha ludens*, across Dietary-restriction  
483 Gradients. *Aging Cell* 7: 470–477.
- 484 Cayol JP (2000) Changes in sexual behavior and life history traits of tephritid species caused by  
485 mass-rearing processes. *Fruit Flies (Tephritidae): Phylogeny and Evolution of Behavior*  
486 (ed. by M Aluja & AL Norrbom) CRC Press, Boca Raton, FL, USA, pp. 843–860.
- 487 Conner JK (2003) Artificial selection: a powerful tool for ecologists. *Ecology* 84:1650–1660.
- 488 Domínguez J, Artiaga-López T, Solís E & Hernández E (2010) Métodos de colonización y cría  
489 masiva. *Moscas de la fruta: Fundamentos y Procedimientos para su Manejo* (ed. by P  
490 Montoya, J Toledo & E Hernández) S y G Editores, Distrito Federal, México, pp. 259–276.
- 491 Ekesi S, Nderitu PW & Chang CL (2007) Adaptation to and small-scale rearing of invasive fruit  
492 fly *Bactrocera invadens* (Diptera: Tephritidae) on artificial diet. *Annals of the*  
493 *Entomological Society of America* 100: 562–567.

494 Enkerlin W (2005) Impact of fruit fly control programmes using sterile insect technique. Sterile  
495 Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management (ed.  
496 by VA Dyck, J Hendrichs & AS Robinson) Springer, Dordrecht, The Netherlands, pp. 651–  
497 676.

498 Falconer DS & Mackay TFC (1996) Introduction to quantitative genetics. Ed.4 Longman Green)  
499 Essex, UK.

500 FAO/IAEA/USDA (2003) Manual for Product Quality Control and Shipping Procedures for  
501 Sterile Mass-Reared Tephritid Fruit Flies. Version 5.0. IAEA, Vienna. 85 pp.

502 Fay MP, Shaw PA (2010) Exact and Asymptotic Weighted Log rank Tests for Interval Censored  
503 Data: The interval R Package. Journal of Statistical Software, 36:1–34.URL  
504 <http://www.jstatsoft.org/v36/i02/>.

505 Gentleman R & Vandal A (2016). Icats: NPMLE for Censored and Truncated Data. R package  
506 version 1.44.0.

507 Gilchrist AS & Meats AW (2014) An evaluation of outcrossing to improve mass-reared strains of  
508 the Queensland fruit fly *Bactrocera tryoni*. International Journal of Tropical Insect Science  
509 34: S35–S44.

510 Greany PD, Szentesi A (1979) Oviposition behavior of laboratory-reared and wild caribbean fruit  
511 flies (*Anastrepha suspense*: Diptera: tephritidae): II. Selected physical influences.  
512 Entomologia Experimentalis et Applicata 26: 239–244.

513 Hendrichs J, Robinson AS, Cayol JP, Enkerlin W (2002) Medfly areawide sterile insect technique  
514 programmes for prevention, suppression or eradication: the importance of mating behavior  
515 studies. Florida Entomologist 85:1–13.



- 516 Hernández E, Toledo J, Artiaga-López T. & Flores S (2009) Demographic changes in *Anastrepha*  
517 *obliqua* (Diptera: Tephritidae) throughout the laboratory colonization process. Journal of  
518 Economic Entomology 102: 542–551.
- 519 Harris DJ, Wood RJ & Bailey SER (1986) Selection for fast and slow mating lines in the Medfly  
520 and analysis of elements of courtship behavior. Pest control: Operations and Systems  
521 Analysis in Fruit Fly Management (ed. by M Mangel, JR Carey & RE Plant) NATO ASI  
522 Series. Series G: Ecological Sciences, Springer-Verlag. Berlin, pp 465.
- 523 Jalvingh KM, Chang PL, Nuzhdin SV & Wertheim B (2014) Genomic changes under rapid  
524 evolution: Selection for parasitoid resistance. Proceedings of the Royal Society B 281: 1–  
525 10.
- 526 Knipling EF (1955) Possibilities of insect control or eradication through the use of sexually sterile  
527 males. Journal of Economic Entomology. 48: 459–462.
- 528 Lance DR, & McInnis DO (2005) Biological basis of the sterile insect technique. Sterile Insect  
529 Technique: Principles and Practice in Area-Wide Integrated Pest Management (ed. by VA  
530 Dyck, J Hendrichs & AS Robinson) Springer, Dordrecht, The Netherlands pp. 69 – 94.
- 531 Lance DR, McInnis DO, Rendón P & Jackson CG (2000) Courtship among sterile and wild  
532 *Ceratitis capitata* (Diptera: Tephritidae) in field cages in Hawaii and Guatemala. Annals  
533 of the Entomological Society of America 93: 1179–1185.
- 534 Liedo P, Carey JR, Celedonio H & Guillen J (1992) Size specific demography of three species of  
535 *Anastrepha* fruit flies. Entomologia Experimentalis et Applicata, 63: 135–142.
- 536 Liedo P & Carey JR (1996) Demography of fruit flies and implications to action programs. Fruit  
537 Fly Pests: A World Assessment of Their Biology and Management (ed. by BA McPherson  
538 & G Steck) St. Lucie Press, Delray Beach, Florida, U.S.A, pp. 299-308.

539 Liedo P, Enkerlin W & Hendrichs J (2010) Fundamentos de la Técnica del Insecto Estéril. Moscas  
540 de la Fruta: Fundamentos y Procedimientos para su Manejo (ed. by P Montoya, J Toledo  
541 & E Hernández) S y G Editores, México, D.F., pp 243-256.

542 Liedo P & Toledo J (2007). Ecología de poblaciones y manejo integrado de las moscas de la fruta  
543 en el Soconusco, Chiapas, México. Moscas de la Fruta en Latinoamérica (Díptera:  
544 Tephritidae): Diversidad, Biología y Manejo (ed. by V Hernández- Ortiz), S y G Editores,  
545 México, D.F., pp. 133-144.

546 Lux SA, Munyiri FN, Vilardi JC, Liedo P, Economopoulos A, Hanson O, Quilici S, Gaggi K,  
547 Cayol JP & Rendon P. (2007) Consistency in courtship pattern among populations of  
548 medfly (Diptera: Tephritidae): comparisons among wild strains and strains mass reared  
549 for SIT operations. Florida Entomologist 85: 113 – 125.

550 Lux SA, Vilardi JC, Liedo P, Gaggi K, Calcagno CE, Munyiri FN, Vera MT, Manso F (2007)  
551 Effects of irradiation on the courtship behavior of medfly (Diptera: Tephritidae) mass  
552 reared for the sterile Insect technique. Florida Entomologist 85: 102– 112.

553 Maathuis M (2013). MLEcens: Computation of the MLE for bivariate (interval) censored data. R  
554 package version 0.1-4. <https://CRAN.R-project.org/package=MLEcens>

555 Matsuyama T & Kuba H (2009) Mating time and call frequency of males between mass-reared  
556 and wild strains of melon fly, *Bactrocera cucurbitae* (Coquillett) (Diptera: Tephritidae).  
557 Applied Entomology and Zoology 44: 309–314.

558 Miyatake T (1995) Two-way artificial selection for developmental period in *Bactrocera*  
559 *cucurbitae* (Diptera: Tephritidae). Annals of the Entomological Society of America  
560 88:848–855.

561 Miyatake T (1998) Genetic changes of life history and behavioral traits during mass-rearing in the  
562 melon fly, *Bactrocera cucurbitae* (Diptera: Tephritidae). *Researches on Population*  
563 *Ecology* 40: 301–310.

564 McInnis DO, Lance DR & Jackson CG (1996) Behavioral resistance to the sterile insect release  
565 technique by the Mediterranean fruit fly (Diptera: Tephritidae) in Hawaii. *Annals of the*  
566 *Entomological Society of America* 89: 739–744.

567 McInnis DO, Shelly TE & Komatsu J (2002) Improving male mating competitiveness and survival  
568 in the field for medfly, *Ceratitidis capitata* (Diptera: Tephritidae) SIT programs. *Genetica*  
569 116: 117–124.

570 Meats A, Holmes HM & Kelly GL (2004) Laboratory adaptation of *Bactrocera tryoni*  
571 (Diptera:Tephritidae) decreases mating age and increases protein consumption and number  
572 of eggs produced per milligram of protein. *Bulletin of Entomological Research* 94: 517–  
573 524.

574 Meza JS, Arredondo J, Orozco D & Perez-Staples D (2014) Disparity in sexual behaviour between  
575 wild and mass-reared Mexican fruit flies. *Physiological Entomology* 39: 263–270

576 Pereira R, Yuval B, Liedo P, Teal PEA, & Shelly TE (2013) Improving sterile male performance  
577 in support of programmes integrating the sterile insect technique. *Journal of Applied*  
578 *Entomology*, 137: 178-190.

579 Perez-Staples D, Martinez-Hernández G & Aluja M (2010) Male age and experience increases  
580 mating success but not female fitness in the Mexican fruit fly. *Ethology* 116: 778–786.

581 Pinheiro J, Bates D, DebRoy S, Sarkar D & R Core Team (2016). nlme: Linear and Nonlinear  
582 Mixed Effects Models. R package version 3.1-127, <URL:[http://CRAN.R-](http://CRAN.R-project.org/package=nlme)  
583 [project.org/package=nlme](http://CRAN.R-project.org/package=nlme)>.

584 Quintero JL, Meza-Hernández JS & Cruz-López L (2009) Biología y comportamiento sexual del  
585 mutante ojos amarillos de *Anastrepha ludens* (Diptera: Tephritidae). Acta Zoológica  
586 Mexicana 25: 9-20.

587 Quintero-Fong L, Toledo J, Ruiz L, Rendon P, Orozco-Dávila D, Cruz L & Liedo P. (2016)  
588 Selection by mating competitiveness improves the performance of *Anastrepha ludens*  
589 males of the genetic sexing strain Tapachula-7. Bulletin of Entomological Research.  
590 doi:10.1017/S0007485316000316.

591 R Core Team (2014). R: A language and environment for statistical computing. R Foundation for  
592 Statistical Computing, Vienna, Austria. URL <https://www.r-project.org>.

593 Robinson AS, & Hendrichs J (2005) Prospects for the future development and application of the  
594 sterile insect technique. Sterile Insect Technique: Principles and Practice in Area-Wide  
595 Integrated Pest Management (ed. by VA Dyck, J Hendrichs & AS Robinson) Springer,  
596 Dordrecht, The Netherlands, pp. 727–760.

597 Rull J, Brunel O & Mendez ME (2005) Mass rearing history negatively affects mating success of  
598 male *Anastrepha ludens* (Diptera:Tephritidae) reared for the sterile insect technique.  
599 Journal of Economic Entomology 100: 1510–1516.

600 Rull J, & Barreda-Landa A (2007) Colonization of a hybrid strain to restore male *Anastrepha*  
601 *ludens* (Diptera: Tephritidae) mating competitiveness for sterile insect technique programs.  
602 Journal of Economic Entomology 100: 752–758.

603 Rull J, Lasa R, Rodriguez C, Ortega R & Velazquez OE et al. (2015) Artificial selection, pre-  
604 release diet, and gut symbiont inoculation effects on sterile male longevity for area-wide  
605 fruit-fly management. Entomologia Experimentalis et Applicata 157: 325–333.

- 606 Schutze MK, Dammalage T, Jessup A, Vreysen MJB, & Wornoayporn V et al. (2015) Effects of  
607 laboratory colonization on *Bactrocera dorsalis* (Diptera, Tephritidae) mating behaviour:  
608 ‘what a difference a year makes’. *ZooKeys* 540: 369:383.
- 609 Sørensen JG, Addison MF & Terblanche JS (2012) Mass-rearing of insects for pest management:  
610 challenges, synergies and advances from evolutionary physiology. *Crop Protection* 38: 87–  
611 94.
- 612 Thomas DB (2003) Reproductive phenology of the Mexican Fruit Fly, *Anastrepha ludens* (Loew)  
613 (Diptera: Tephritidae) in the Sierra Madre Oriental, Northern Mexico. *Neotropical*  
614 *Entomology* 32: 385–397.
- 615 Vargas RI & Carey JR (1989) Comparison of demographic parameters for wild and laboratory  
616 adapted Mediterranean fruit fly (Diptera: Tephritidae). *Annals of the Entomological*  
617 *Society of America* 82: 55–59.
- 618 Vargas RI, Walsh WA, Kanehisa D, Stark JD & Nishida T (2000) Comparative demography of  
619 three Hawaiian fruit flies (Diptera: Tephritidae) at alternating temperatures. *Annals of the*  
620 *Entomological Society of America* 93: 75–81.
- 621 Vera MT, Cladera JL, Calcagno G, Vilardi JC, Mcinnis DO & field working group (2003)  
622 Remating of Wild *Ceratitis capitata* (Diptera: Tephritidae) Females in Field Cages. *Annals*  
623 *of the Entomological Society of America* 96: 563–570 (2003)
- 624 Zar, JH. (2010). *Biostatistical Analysis* (5th ed.). Englewood Cliffs, NJ: Prentice-Hall, pp 944.

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627 **Figure legends**

628 Figure 1. Flow diagram of the process showing selection events starting from the laboratory  
629 parental males through 3 generations (F1, F2 and F3). The initial number of males tested and the  
630 number and percentage of males selected is shown.

631  
632 Figure 2. Mating competitiveness. Proportion of matings in field cage tests by AS, BS, L and W  
633 males in four consecutive generations (A = F1, B = F2, C = F3 and D = F4). Different letters  
634 indicate significant differences among treatments within each generation according to Tukey HSD  
635 test with  $P < 0.05$ .

636  
637 Figure 3. Wild female mating percentage (light gray) with males from different treatments (W, L,  
638 AS and BS), and rematings (dark gray) with L virgin males during four consecutive generations  
639 (A = F1, B = F2, C = F3, and D = F4). Different letters within each generation indicate significant  
640 differences among treatments according to Tukey HSD test with  $P < 0.05$

641  
642 Figure 4. Number of surviving individuals in each stage (larva, pupa, adult) in each treatment (L,  
643 AS, BS) per generations (A = F1, B = F2, C = F3). Each point represents a replicate and the solid  
644 line represents the mean.

645  
646 Figure 5. Survival of females (left) and males (right) from three mass-rearing colonies (L, AS  
647 and BS) and the W strain during three consecutive generations (Females: A = F1, B = F2 and C  
648 = F3; Males: D = F1, E = F2 and F = F3). The same data from the wild strain was used as a  
649 control for the three generations.

650

651 Figure 6. Gross fecundity. Mean number of eggs laid per female per day in each of the three  
652 mass-reared colonies (L, AS and BS) and the W strain during three consecutive generations (A =  
653 F1, B = F2 and C = F3). The same data from the wild strain was used as a control for the three  
654 generations.

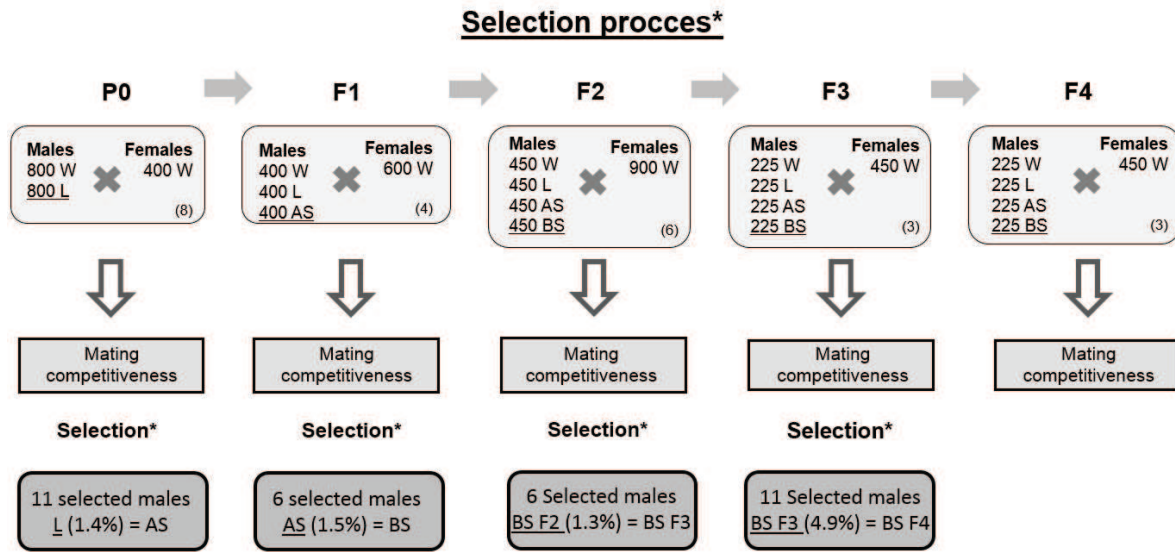
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656 Figure 7. Average of the proportion of hatched eggs (fertility) in each treatment (L, AS, BS)  
657 during 3 consecutive generations (A = F1, B = F2, C = F3).

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660 Figure 1



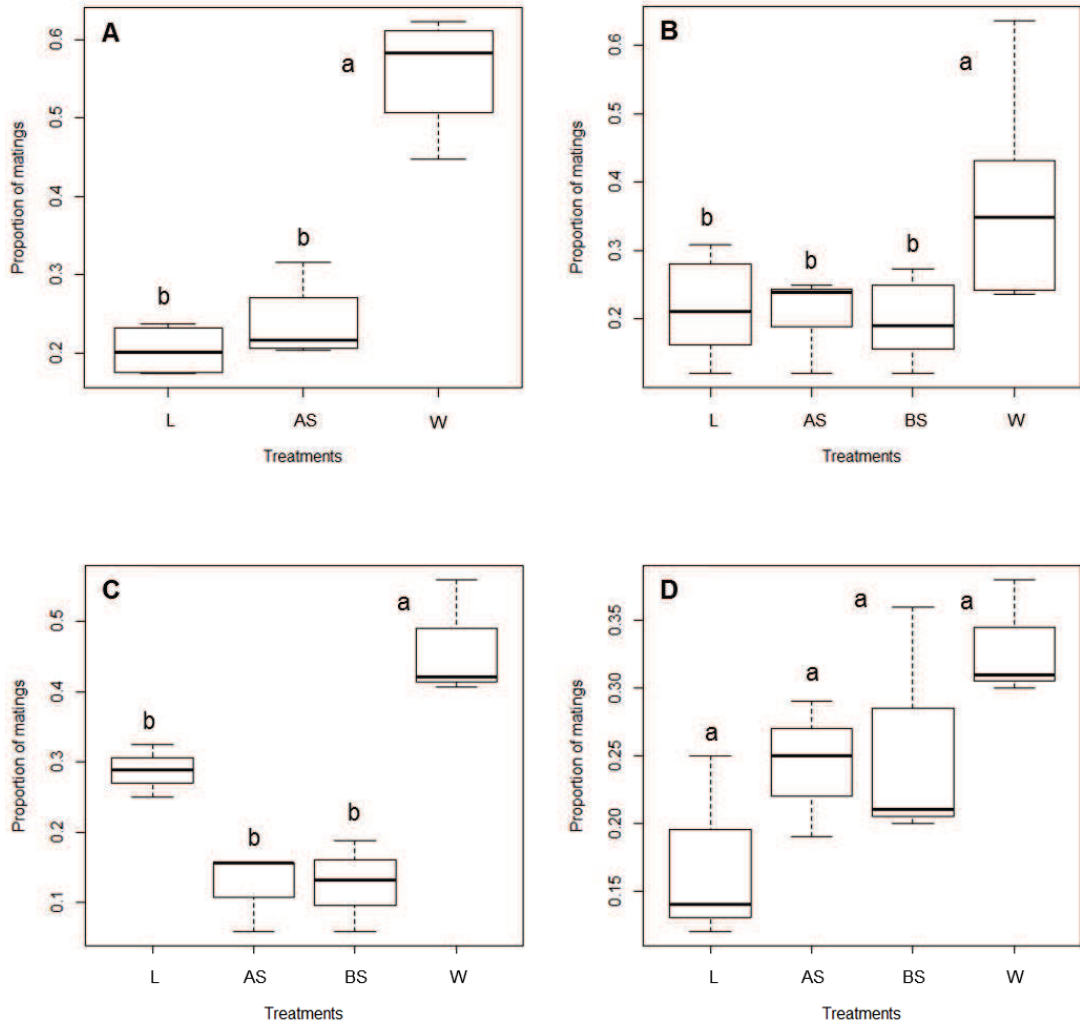
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664 Figure 2



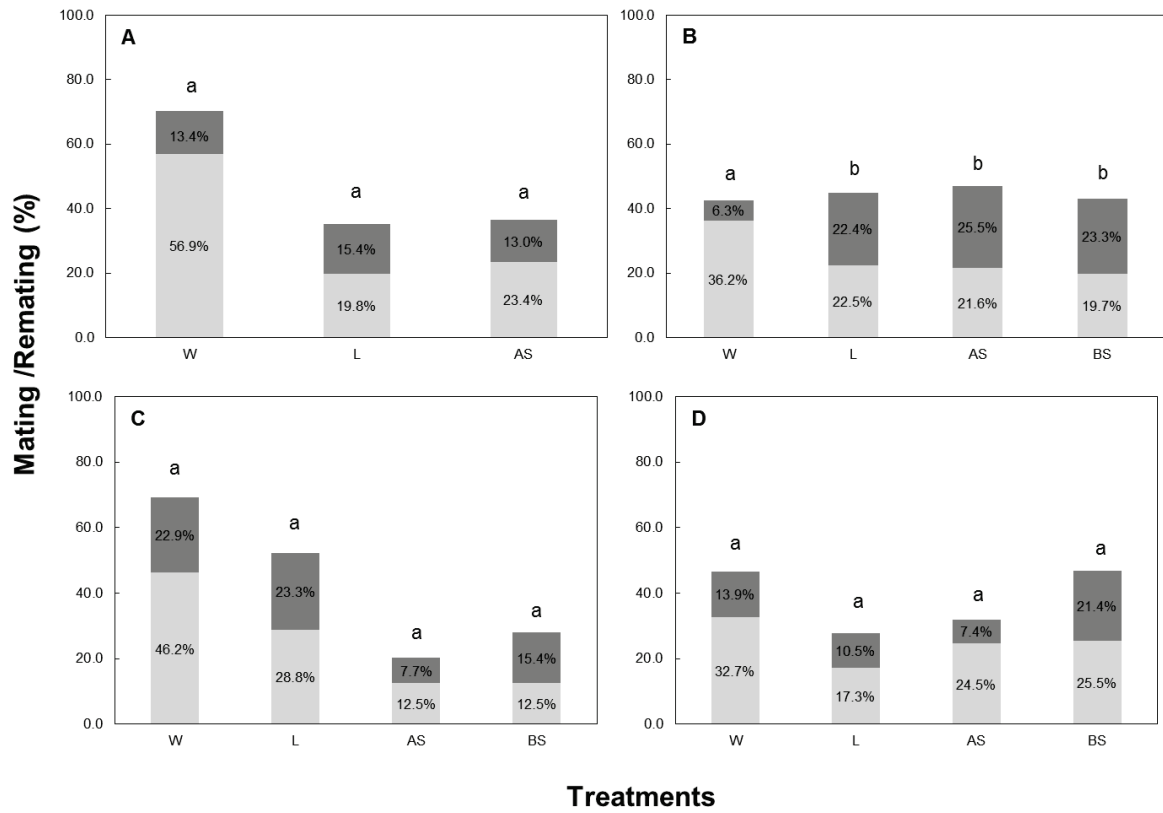
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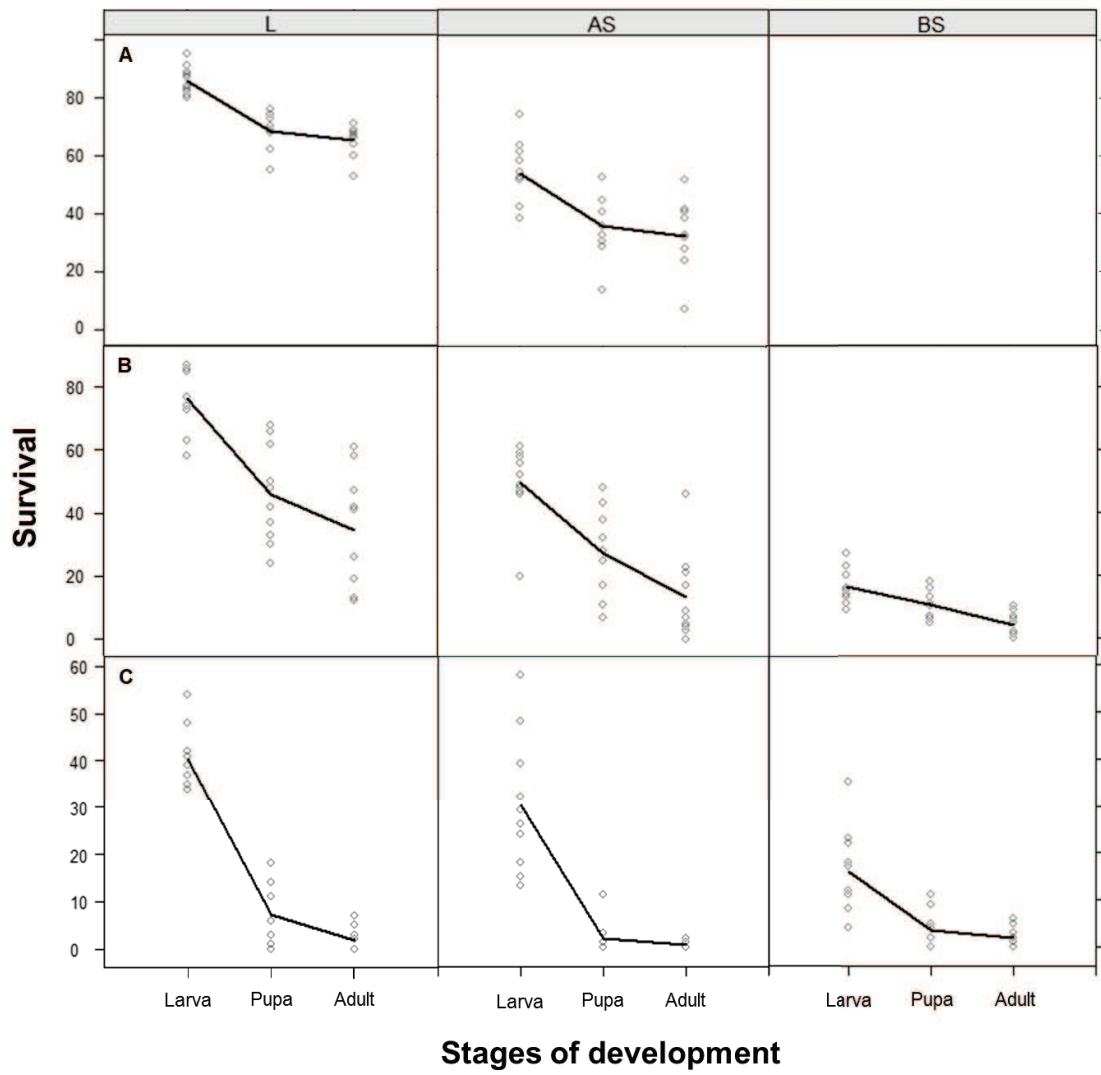
669 Figure 3



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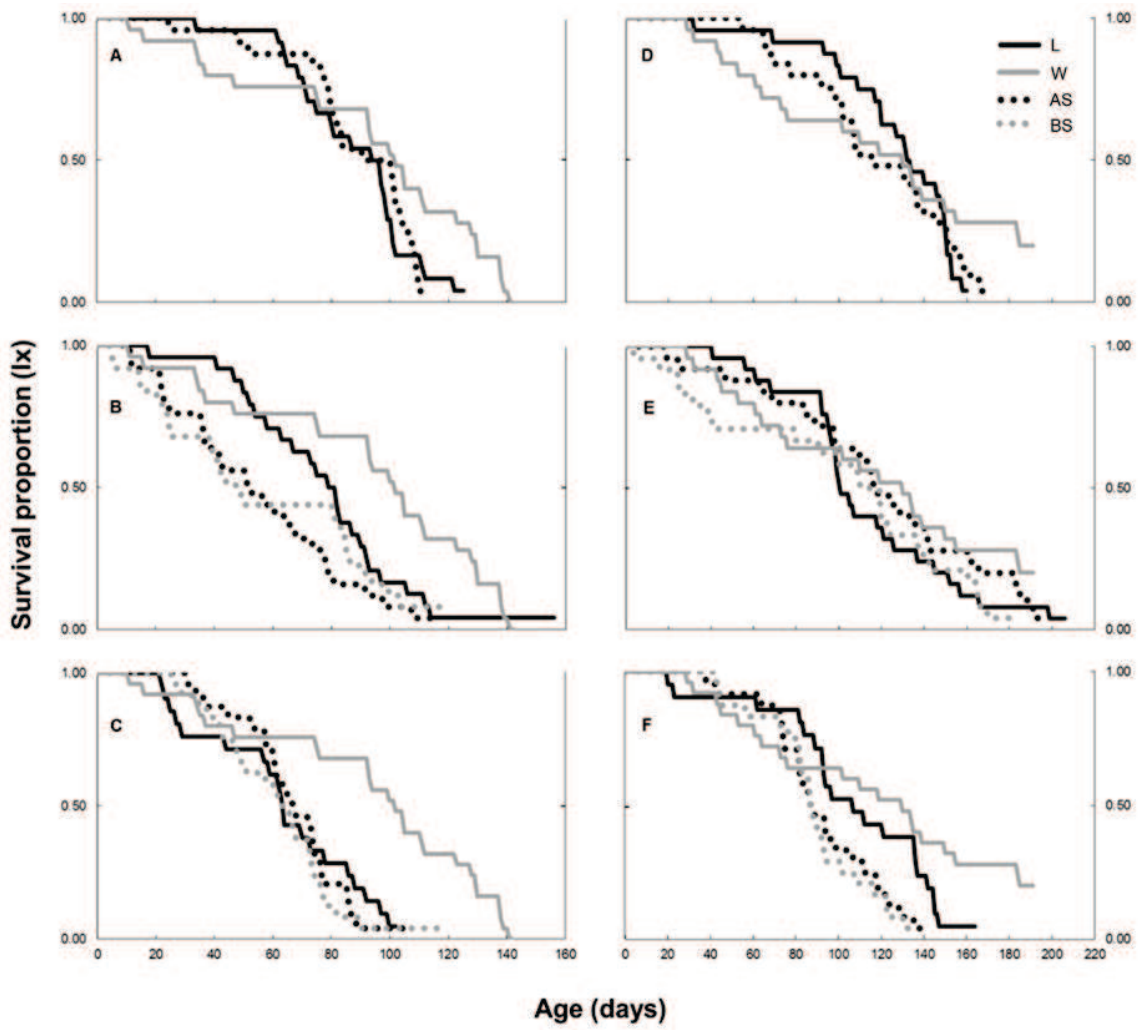


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676 Figure 5



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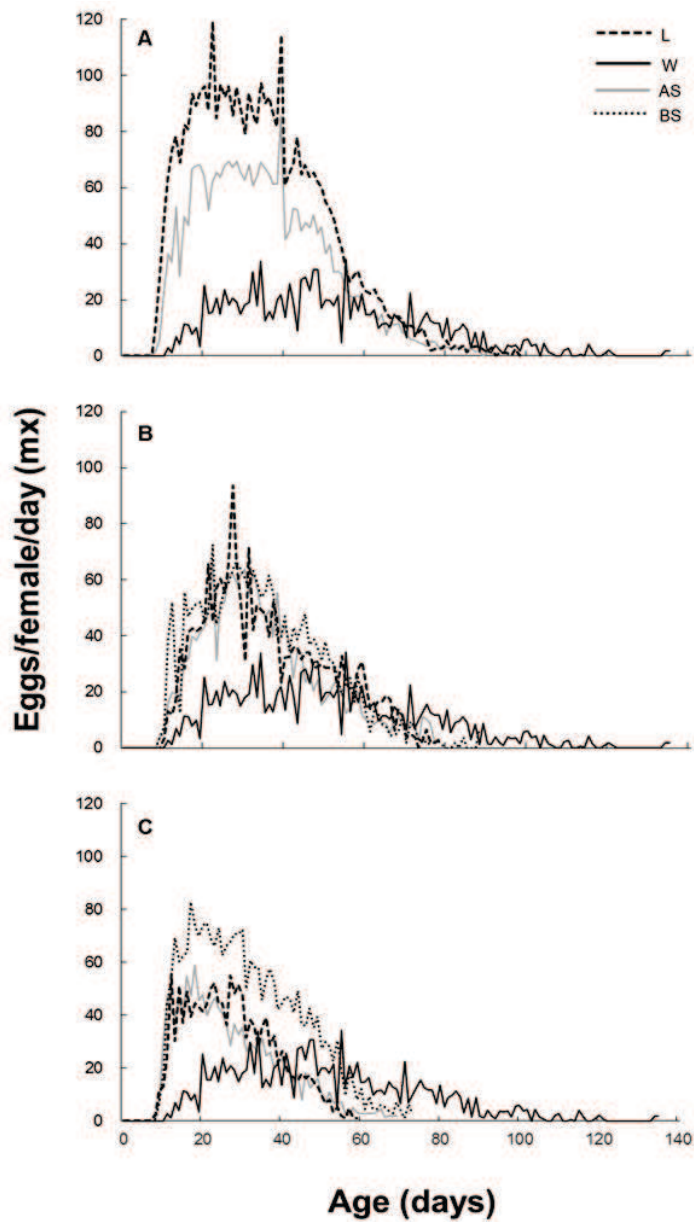
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686 Figure 6

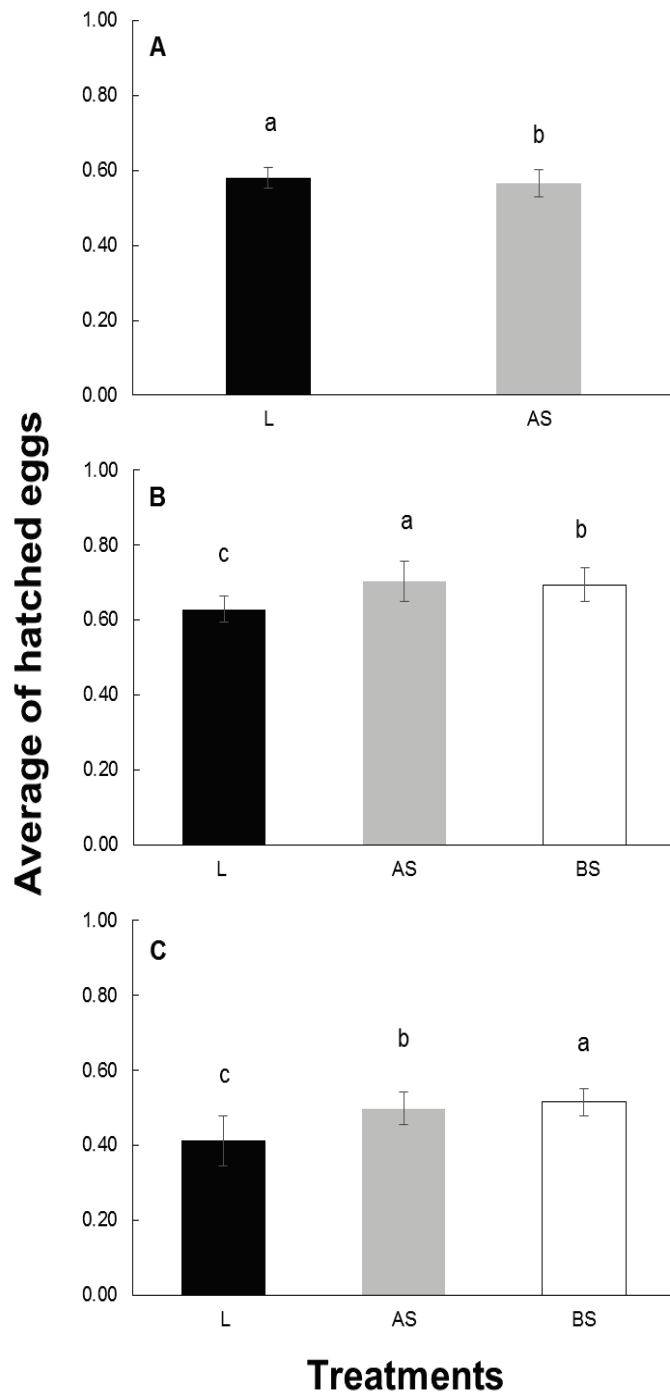


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690 Figure 7



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695 Table 1. Mean ( $\pm$ SE) mating latency and copula duration (minutes) of wild flies mated with males  
 696 from different treatments in field cage tests. Different letters in each column indicate significant  
 697 differences according to Tukey HSD test with  $P < 0.05$ .

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Treatments males	Mating latency				Copulation duration			
	F1	F2	F3	F4	F1	F2	F3	F4
Wild	84 $\pm$ 2 a	91 $\pm$ 3 b	83 $\pm$ 3 a	52 $\pm$ 2 a	37 $\pm$ 3 a	60 $\pm$ 4 b	36 $\pm$ 5 a	39 $\pm$ 5 a
Laboratory	78 $\pm$ 5 a	72 $\pm$ 4 a	76 $\pm$ 5 a	46 $\pm$ 4 a	33 $\pm$ 4 a	42 $\pm$ 4 a	56 $\pm$ 10 a	40 $\pm$ 8 a
Alpha Selection	84 $\pm$ 4 a	72 $\pm$ 4 a	70 $\pm$ 6 a	49 $\pm$ 3 a	42 $\pm$ 5 a	33 $\pm$ 4 a	36 $\pm$ 9 a	37 $\pm$ 6 a
Beta Selection	—	81 $\pm$ 5 ab	89 $\pm$ 5 a	47 $\pm$ 3 a	—	42 $\pm$ 5 a	57 $\pm$ 17 a	44 $\pm$ 6 a

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719 Table 2. Demographic parameters of male and female *Anastrepha ludens* flies under different  
 720 selection methods for mass-rearing.

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Parameter	Wild	Laboratory			Alpha Selection			Beta Selection	
		F1	F2	F3	F1	F2	F3	F2	F3
e <sub>0</sub> male (days)	118.66	126.38	112.98	105.69	117.90	119.34	92.17	100.38	88.75
e <sub>0</sub> female (days)	92.02	87.79	77.50	62.64	88.25	55.26	66.46	57.26	61.29
Gross fecundity (mx)	1256.67	3984.53	2110.73	1426.01	2736.22	1938.37	1373.43	2295.36	2550.70
Net fecundity (l <sub>x</sub> m <sub>x</sub> )	994.72	3860.54	1909.17	1231.43	2605.46	1360.80	1307.67	1510.28	2257.63
Daily eggs (net)	10.80	43.97	24.63	19.66	29.52	24.63	19.68	26.38	36.83
Daily eggs (gross)	13.65	45.39	27.24	22.76	31.01	35.08	20.67	40.09	41.62



### III. Discusión General y Conclusiones

Los resultados de nuestro trabajo proporcionan nuevas evidencias que muestran la complejidad del proceso de selección fundamentado en el éxito de apareamiento, destacando que el éxito reproductivo se encuentra asociado a inherentes costos o compromisos biológicos (“trade-off”).

De acuerdo con nuestro objetivo de comparar la competitividad sexual de machos de *A. ludens* provenientes de un solo evento de selección (SA) con machos provenientes de selección continúa a lo largo de cuatro generaciones (SB), los resultados de las pruebas de competitividad sexual indicaron que no existió diferencia entre las dos variantes del proceso de selección (SA y SB). Al comparar el desempeño de los machos provenientes de colonias seleccionadas con el testigo de laboratorio (L), en la cuarta generación, se pudo apreciar que aunque no hubo diferencia significativa, sí hubo una tendencia a una mayor competitividad.

Por otro lado, los rasgos de historia de vida, en particular la fecundidad y fertilidad, mostraron que el efecto de la selección continua (SB) resultó en aumentos que son favorables para la cría masiva.

Estudios previos de selección artificial fundamentados en el desempeño sexual de los machos (Bosa et al., 2016; Quintero et al., 2016), aunado a nuestros resultados permiten especular acerca de las principales causas que pudiesen haber enmascarado el potencial del proceso de selección. Destaca los elevados porcentajes de presión de selección, los cuales fueron >95%, en adición a la variabilidad y al decremento del tamaño poblacional (800 en parentales hasta 225 en las últimas generaciones) lo que pudiese haber resultado en recombinación y

deriva genética, contrarrestando el efecto de la selección (Falconer y Mackay 1996, Conner 2003, Jalvingh et al. 2014).

Otra razón pudiese ser las altas relaciones de hembras de laboratorio: machos exitosos, que fueron utilizadas para garantizar la descendencia de los machos seleccionados, lo que resultó en una mayor variación genética que pudo disminuir la frecuencia de los genes asociados a la competitividad sexual de los machos seleccionados.

Los resultados obtenidos en la sobrevivencia de los estados inmaduros, particularmente la sobrevivencia o eclosión de huevos, constituyen un reflejo de las altas proporciones de hembras de laboratorio: machos exitosos, debido a que la gran cantidad de hembras utilizadas y el número reducido de machos, sobretodo en el tratamiento de selección continua (SB), pudieron provocar que muchas hembras no se hayan apareado y por lo tanto muchos huevos no fueran fertilizados.

La latencia a obtener una cópula tendió a ser más larga en las moscas silvestres (W) y más corta en los machos de laboratorio (L). Los machos de las líneas seleccionadas (SA y SB) tuvieron una latencia a la cópula intermedia, aunque las diferencias no fueron significativas. Esto puede deberse a que la latencia a la cópula es determinada por las hembras, que en nuestro caso siempre fueron silvestres (Briceño y Eberhard 2002). Los machos de laboratorio generalmente iniciaban el llamado y comportamiento de cortejo más temprano que los silvestres, pero esto no resultó en mayores copula (Bosa et al. 2016, Meza et al. 2014). Respecto a la duración de la cópula, no se encontró una asociación entre la duración de la cópula y el tratamiento, y esto pudo deberse a que este parámetro está regulado

principalmente por las hembras y todas las hembras utilizadas fueron de origen silvestre (Pérez-Staples et al. 2010).

No se encontraron diferencias significativas en la habilidad de los machos seleccionados en inhibir la receptividad sexual de las hembras con las cuales copularon, ya que no existió una relación entre el tipo de macho de la primera cópula con la propensión de la hembra a copular nuevamente. Esto sugiere que los machos de cría masiva tienen el mismo potencial que los silvestres para inhibir el reapareamiento, lo cual resulta favorable para la técnica del insecto estéril, aunque habría que considerar el efecto de la irradiación (Gavriel et al. 2009, Pérez-Staples et al. 2012).

Las diferencias en la sobrevivencia de adultos silvestres y laboratorio fueron las típicas reportadas en otros estudios (Foote y Carey 1987, Vargas & Carey 1989, Sivinsky 1993, Liedo y Carey 1996, Liedo et al. 2007). En los primeros 20 a 30 días de edad las moscas de laboratorio registraron una mayor sobrevivencia que las silvestres, posteriormente hubo una acelerada mortalidad en las de laboratorio, reflejándose en una menor longevidad que las moscas silvestres. Entre las líneas seleccionadas y la colonia de laboratorio no seleccionada (testigo) las diferencias fueron menores y en algunos casos no significativas.

Aunque los machos silvestres fueron más competitivos respecto a los de las colonias de laboratorio (L, SA, SB), los rasgos de historia de vida sugieren que la competitividad sexual puede estar comprometida con otros atributos biológicos, como la sobrevivencia o la reproducción (Carey et al. 2005). La comprensión de estos rasgos poligénicos y sus compromisos (trade-offs) representan una ventana de opciones para mejorar el desempeño del macho estéril.

## Conclusiones:

- Se observaron diferencias consistentes entre moscas silvestres y de laboratorio en la competitividad sexual, sobrevivencia de adultos y fecundidad. No se observaron diferencias o fueron menos consistentes en latencia a la cópula, duración de la cópula e inhibición al apareamiento.
- Las diferencias entre moscas de laboratorio no seleccionadas y seleccionados fueron menores o no significativas.
- Los machos de laboratorio mostraron mayor sobrevivencia de inmaduros, mientras que los tratamientos de selección beta y selección alfa mostraron una mayor fecundidad y fertilidad.
- Los procesos de selección no indujeron una mayor competitividad sexual en los machos.
- La mayor fecundidad y fertilidad de los tratamientos con selección puede contribuir a mejorar la cría masiva.
- La menor sobrevivencia, particularmente de hembras de cría masiva, puede explicarse como el costo de la reproducción ya que tuvieron mayores tasas de fecundidad.

#### IV. Literatura citada

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- Arita LH, Kaneshiro KY. 1989. Sexual Selection and Lek Behavior in the Mediterranean Fruit Fly, *Ceratitis capitata* (Diptera: Tephritidae). *Annu. Rev. Entomol.* 43: 135–143.
- Aluja M, Mangan RL. 2008. Fruit fly (Diptera: Tephritidae) host status determination: critical conceptual, methodological, and regulatory considerations. *Annu. Rev. Entomol.* 53: 473–502.
- Aluja M, Arredondo J, Díaz-Fleischer F, Birke A, Rull J, Niogret J, Epsky N. 2014. Susceptibility of 15 mango (Sapindales: Anacardiaceae) cultivars to the attack by *Anastrepha ludens* and *Anastrepha obliqua* (Diptera: Tephritidae) and the role of underdeveloped fruit as pest reservoirs: management implications. *J. Econ. Entomol.* 107: 375-388.
- Hernández-Ortiz V, Aluja M. 1993. Listado de especies del género neotropical *Anastrepha* (Diptera: Tephritidae) con notas sobre su distribución y plantas hospederas. *Folia Entomol. Mex.* 88: 89-105.
- Benelli G, Canale A, Bonsignori G, Ragni G, Stefanini C, Raspi A. 2012. Male wing vibration in the mating behavior of the olive fruit fly *Bactrocera oleae* (Rossi) (Diptera: Tephritidae). *J. Insect Behav.* 25:590–603.
- Benelli G, Giunti G, Canale A, Messing RH. 2014. Lek dynamics and cues evoking mating behavior in tephritid flies infesting soft fruits: implications for behavior-based control tools. *Appl. Entomol. Zool.* 49: 363–373.
- Boake CRB. 1996. Sexual selection in relation to strategies pest-management. *Annu. Rev. Entomol.* 41: 211-29.

- Bosa CF, Cruz-López L, Zepeda-Cisneros CS, Valle-Mora J, Guillén-Navarro K, Liedo P. 2016. Sexual behavior and male volatile compounds in wild and mass-reared strains of the Mexican fruit fly *Anastrepha ludens* (Diptera: Tephritidae) held under different colony management regimes. *Insect Science*. 23: 106-116.
- Briceño RD, Eberhard WG. 2002. Decisions during courtship by male and female medflies (Diptera: Tephritidae): correlated changes in male and female acceptance criteria in mass-reared flies. *Fla. Entomol.* 85: 14–31.
- Burk T. 1981. Signaling and sex in acalyptrate flies. *Fla. Entomol.* 64: 30–43.
- Calkins CO. 1984. The importance of understanding fruit fly mating behavior in sterile male release programs (Diptera: Tephritidae). *Folia Entomol. Mex.* 61: 205–213.
- Carey JR, Liedo P, Müller HG, Wang JL, Senturk D, Harshman L. 2005. Biodemography of a long-lived tephritid: Reproduction and longevity in a large cohort of Mexican fruit flies, *Anastrepha ludens*. *Exp. Geront.* 40:793–800.
- Cayol JP. 2000. Changes in sexual behavior and life history traits of tephritid species caused by mass-rearing processes. *En: Aluja M, Norrbom AAL. (eds.). Fruit Flies (Tephritidae): Phylogeny and Evolution of Behavior.* Boca Raton, Florida. CRC Press. pp. 843-860.
- Conner JK. 2003. Artificial selection: a powerful tool for ecologists. *Ecology.* 84: 1650–1660.
- Eberhard WG. 1996. *Female control: Sexual selection by cryptic female choice.* NJ, U.S.A. Princeton University Press.

- Enkerlin W. 2005. Impact of fruit fly control programmes using sterile insect technique. *En*: Dyck VA, Hendrichs J, Robinson AS (eds.). *Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management*. Springer, Dordrecht, The Netherlands. pp. 651–676.
- Falconer DS, Mackay TFC. 1996. *Introduction to quantitative genetics*. 4a. Ed. Longman.
- Foote D, Carey JR. 1987. Comparative demography of a laboratory and a wild strain of the oriental fruit fly, *Dacus dorsalis*. *Entomol. Exp. Appl.* 44: 263-268.
- Gavriel S, Gazit Y, Yuval B. 2009. Remating by female Mediterranean fruit flies (*Ceratitis capitata*, Diptera: Tephritidae): Temporal patterns and modulation by male condition. *J. Insect Physiol.* 55: 637–642.
- Gilchrist AS, Meats AW. 2014. An evaluation of outcrossing to improve mass-reared strains of the Queensland fruit fly *Bactrocera tryoni*. *Int. J. Trop. Insect Sci.* 34: S35–S44.
- Gutiérrez JM. 2010. El programa de moscas de la fruta en México. *En*: Montoya P, Toledo J, Hernández E. (eds.). *Moscas de la fruta: Fundamentos y Procedimientos para su Manejo*. S y G Editores. México, D. F. pp: 3-10.
- Gutiérrez JM, Santiago G. 2008. Situación actual de la campaña nacional contra moscas de la fruta en México. *En*: Montoya P, Díaz F, Flores S. *Memorias de la Séptima Reunión del Grupo de Trabajo en Moscas de la Fruta del Hemisferio Occidental*. Noviembre 2 a 7, Mazatlán, Sinaloa, México. pp. 11-13.
- Harris DJ, Wood RJ, Bailey SER. 1986. Selection for fast and slow mating lines in the Medfly and analysis of elements of courtship behavior. *En*: Mangel M,

- Carey JR, Plant RE (eds.). Pest control: Operations and Systems Analysis in Fruit Fly Management. NATO ASI Series. Series G: Ecological Sciences. Berlin. Springer-Verlag. pp 465.
- Harris DJ, Wood RJ, Bailey SER. 1988. Two way selection for mating activity in the Mediterranean fruit fly, *Ceratitidis capitata*. Entomol. Exp. & Appl. 47: 239-248.
- Jalvingh KM, Chang PL, Nuzhdin SV, Wertheim B. 2014. Genomic changes under rapid evolution: Selection for parasitoid resistance. Proc. R. Entomol. Soc. B. 281: 1–10.
- Hendrichs J, Robinson A, Cayol JP, Enkerlin W. 2002. Medfly areawide sterile insect technique programmes for prevention, suppression or eradication: the importance of mating behavior studies. Fla. Entomol. 85: 1-13
- Hendrichs MA, Wornoayporn V, Katsoyannos B, Hendrichs J. 2007. Quality control method to measure predator evasion in wild and mass-reared Mediterranean fruit flies (Diptera: Tephritidae). Fla. Entomol. 90: 64–70.
- Hernandez-Ortíz V. 2007. Distribución y biogeografía del género *Anastrepha* en México. En: Hernández-Ortiz V, (ed.). Moscas de la Fruta en Latinoamérica, Diversidad, Biología y Manejo. S y G Editores. México, D.F. pp: 53-76
- Hernández-Ortiz V, Guillén-Aguilar J, López L. 2010. Taxonomía e identificación de moscas de la fruta de importancia económica en América. En: Montoya P, Toledo J, Hernández E. (eds.). Moscas de la Fruta: Fundamentos y Procedimientos para su Manejo. México, D.F. S y G Editores. pp: 49-80.
- Knipling EF. 1955. Possibilities of insect control or eradication through the use of sexual sterile males. J. Econ. Entomol. 48: 459-462.



- Lance DR, McInnis DO. 2005. Biological basis of the sterile insect technique. *En: Dyck A, Hendrichs J, Robinson AS. (eds.). Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management. The Netherlands. Spring Verlag. pp. 69-94*
- Liedo P, Carey JR. 1996. Demography of fruit flies and implications to action programs. *En: McPheron BA, Steck G. (eds.). Fruit Fly Pests: A world assessment of their biology and management. Delray Beach, Florida, U.S.A. St. Lucie Press. pp. 299-308*
- Liedo P, Salgado S, Oropeza A, Toledo J. 2007. Improving mating performance of mass-reared sterile Mediterranean fruit flies (Diptera: Tephritidae) through changes in adult holding conditions: Demography and mating competitiveness. *Fla. Entomol. 90: 33–40.*
- Liedo P, Enkerlin W, Hendrichs J. 2010. Fundamentos de la técnica del insecto Estéril. *En: Montoya P, Toledo J, Hernández E. (eds.). Moscas de la Fruta: Fundamentos y Procedimientos para su Manejo. México, D.F. S y G Editores. pp: 243-256*
- Lux SA, Vilardi JC, Liedo P, Gaggi K, Calcagno GE, Munyiri FN. 2002. Effects of irradiation on the courtship behaviour of medfly (Diptera, Tephritidae) mass reared for the sterile insect technique. *Fla. Entomol. 85: 102–112.*
- Meza JS, Arredondo J, Orozco D, Pérez-Staples D. 2014. Disparity in sexual behaviour between wild and mass-reared Mexican fruit flies. *Physiol. Entomol. 39: 263–270.*

- McInnis DO, Rendon P, Komatsu J. 2002. Mating and remating of medflies (Diptera: Tephritidae) in Guatemala: individual fly marking in field cages. Fla. Entomol. 85: 126–137.
- NOM-023-FITO-1995. Por la que se establece la Campaña Nacional Contra Moscas de la Fruta. Diario Oficial de la Federación. 11 de febrero de 1999. México, D. F.
- NOM-075-FITO-1997. Por la que se establecen los requisitos y especificaciones fitosanitarias para la movilización de frutos hospederos de moscas de la fruta. Diario Oficial de la Federación. 16 de marzo de 1998. México, D. F.
- Pereira R, Yuval B, Liedo P, Teal PEA, Shelly TE. 2013. Improving sterile ale performance in support of programmes integrating the sterile insect technique. J. Appl. Entomol. 137: 178-190.
- Pérez-Staples D, Martínez-Hernández MG, Aluja M. 2010. Male age and experience increases mating success but not female fitness in the Mexican fruit fly. Ethol. 116: 778–786.
- Perez-Staples D, Shelly TE, Yuval B. 2012. Female mating failure and the failure of “mating” in sterile insect programs. Entomol. Exp. Appl. 146: 66–78.
- Prokopy RJ, Hendrichs J. 1979. Mating behavior of *Ceratitidis capitata* on a field caged host tree. Ann. Entomol. Soc. Am. 72: 642-648.
- Quintero-Fong L, Toledo J, Ruiz L, Rendon P, Orozco-Dávila D, Cruz L, Liedo P. 2016. Selection by mating competitiveness improves the performance of *Anastrepha ludens* males of the genetic sexing strain Tapachula-7. Bull. Entomol. Res. doi:10.1017/S0007485316000316.

- Reyes J, Santiago G, Hernández P. 2000. The Mexican fruit fly eradication programme. *En: Tan KH. (ed.) Penerbit Universiti Sains Malaysia, Area-Wide Control of Fruit Flies and Other Insect Pests. Pulau Pinang, Malaysia. pp. 377-380.*
- Robinson AS, Hendrichs J. 2005. Prospects for the future development and application of the sterile insect technique. *En: Dyck VA, Hendrichs J, Robinson AS (eds.). Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management. Springer, Dordrecht, The Netherlands. pp. 727–760.*
- Rull J, Reyes J, Enkerlin W. 1996. The Mexican national fruit fly eradication campaign: Largest fruit fly industrial complex in the world. *En: McPheron BA, Steck GJ (eds.). Fruit Fly Pests. A World Assessment of Their Biology and Management. Delray Beach. St. Lucie Press. pp. 561-563.*
- Rull J, Brunel O, Méndez ME. 2005. Mass rearing history negatively affects mating success of male *Anastrepha ludens* (Diptera: Tephritidae) reared for sterile insect technique programs. *J. Econ. Entomol. 95: 1510–1516.*
- Rull J, Barreda-Landa A. 2007. Colonization of a hybrid strain to restore male *Anastrepha ludens* (Diptera: Tephritidae) mating competitiveness for sterile insect technique programs. *J. Econ. Entomol. 100: 752–758.*
- Sivinski J. 1993. Longevity and fecundity in the Caribbean fruit fly (Diptera: Tephritidae): effects of mating, strain and body size. *Fla. Entomol. 76: 635–644.*

- Sørensen JG, Addison MF, Terblanche JS. 2012. Mass-rearing of insects for pest management: challenges, synergies and advances from evolutionary physiology. *Crop Prot.* 38: 87–94.
- Vargas RI, Carey JR. 1989. Comparison of demographic parameters for wild and laboratory-adapted Mediterranean fruit fly (Diptera: Tephritidae). *Ann. Entomol. Soc. Am.* 82: 55-59
- Whittier TS, Kaneshiro KY. 1995. Intersexual selection in the Mediterranean fruit fly: Does female choice enhance fitness? *Evol.* 49: 990.

